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CHEMICAL ABSTRACTS, vol. 92, no. 15, 14th April 1980, page 430, abstract no. 124751q, Columbus, US; A. HINNEN et al.: "Isolation of a yeast gene (HIS4) by transformation of yeast"

(73) Proprietor: **PHILLIPS PETROLEUM COMPANY**
5th and Keeler
Bartlesville Oklahoma 74004(US)

(72) Inventor: **Stroman, David Womack**
2504 SW Centre Rd.
Bartlesville Oklahoma 74006(US)
Inventor: **Cregg, James Michael**
6225 Via Regla
San Diego California 92122(US)
Inventor: **Sperl, George T.**
906 Taylor Drive
Gurnee Illinois 60031(US)
Inventor: **Harpold, Michael Miller**
1341 29th Street
San Diego California 92102(US)

(74) Representative: **Dost, Wolfgang, Dr.rer.nat., Dipl.-Chem. et al**
Patent- und Rechtsanwälte Bardehle-Pagenberg-Dost-Altenburg & Partner Postfach 86 06 20
W-8000 München 86(DE)

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CHEMICAL ABSTRACTS, vol. 97, no. 19, 8th November 1982, page 193, astract no. 157352s, Columbus, Ohio, US; T.F. DONAHUE et al.: "The nucleotide sequence of the HIS4 region of yeast" JCN-UCLA Symp. Mol.: Cellular Biology, 1979, 14 (Eucaroytic Gene Regulation); pp. 43-50	

Description

Background

This invention relates to the field of recombinant DNA technology. In one of its aspects, the invention relates to novel yeast strains. In another aspect, the invention relates to processes for transforming yeast strains with recombinant DNA material.

Up to now, commercial efforts employing recombinant DNA technology for producing various polypeptides have centered on *Escherichia coli* as a host organism. However, in some situations *E. coli* may prove to be unsuitable as a host. For example, *E. coli* contains a number of toxic pyrogenic factors that must be eliminated from any polypeptide useful as a pharmaceutical product. The efficiency with which this purification can be achieved will, of course, vary with the particular polypeptide. In addition, the proteolytic activities of *E. coli* can seriously limit yields of some useful products. These and other considerations have led to increased interest in alternative hosts, in particular, the use of eukaryotic organisms for the production of polypeptide products is appealing.

The availability of means for the production of polypeptide products in eukaryotic systems, e.g., yeast, could provide significant advantages relative to the use of prokaryotic systems such as *E. coli* for the production of polypeptides encoded by recombinant DNA. Yeast has been employed in large scale fermentations for centuries, as compared to the relatively recent advent of large scale *E. coli* fermentations. Yeast can generally be grown to higher cell densities than bacteria and are readily adaptable to continuous fermentation processing. In fact, growth of yeast such as *Pichia pastoris* to ultra-high cell densities, i.e., cell densities in excess of 100 g/L, is disclosed by Wegner in U.S. 4,414,329 (assigned to Phillips Petroleum Co.). Additional advantages of yeast hosts include the fact that many critical functions of the organism, e.g., oxidative phosphorylation, are located within organelles, and hence not exposed to the possible deleterious effects of the organism's production of polypeptides foreign to the wild-type host cells. As a eukaryotic organism, yeast may prove capable of glycosylating expressed polypeptide products where such glycosylation is important to the bioactivity of the polypeptide product. It is also possible that as a eukaryotic organism, yeast will exhibit the same codon preferences as higher organisms, thus tending toward more efficient production of expression products from mammalian genes or from complementary DNA (cDNA) obtained by reverse transcription from, for example, mammalian mRNA.

The development of poorly characterized yeast species as host/vector systems is severely hampered by the lack of knowledge about transformation conditions and suitable vectors. In addition, auxotrophic mutations are often not available, precluding a direct selection for transformants by auxotrophic complementation. If recombinant DNA technology is to fully sustain its promise, new host/vector systems must be devised which facilitate the manipulation of DNA as well as optimize expression of inserted DNA sequences so that the desired polypeptide products can be prepared under controlled conditions and in high yield.

Objects of the Invention

An object of our invention, therefore, is the transformation of yeast of the species *Pichia pastoris*.

Another object of our invention is a host of the species *Pichia pastoris* for transformation with recombinant DNA material.

These and other objects of the invention will become apparent from the disclosure and claims herein provided.

Statement of the Invention

In accordance with the present invention, we have developed a process for the transformation of yeast cells of the species *Pichia pastoris*. By the practice of the transformation process of the present invention, DNA sequences can be introduced into host cells of the species *Pichia pastoris* allowing *Pichia pastoris* to be employed as a host system for the production of polypeptide product in yeast.

Further, in accordance with the present invention, novel strains of microorganisms of the species *Pichia pastoris* are provided. These novel strains are useful as hosts for the introduction of recombinant DNA material into yeast.

Brief Description of the Drawings

Figure 1 is a restriction map of plasmid pYA2.

Figure 2 is a restriction map of plasmid YEp13.
Figure 3 is a restriction map of plasmid pYA4.
Figure 4 is a restriction map of plasmid pYJ30.
Figure 5 is a restriction map of plasmid pYJ32.
Figure 6 is a restriction map of plasmid pSAOH5.

The following abbreviations are used throughout this application to represent the restriction enzymes employed:

Abbreviation	Restriction Enzyme
B	*Bam*HI
B_2	*Bgl*II
H_3	*Hin*dIII
Nr	*Nru*I
Ps	*Pst*I
R_1	*Eco*RI
R_5	*Eco*RV
S	*Sal*I
Sm	*Sma*I
Sp	*Sph*I
S_3	*Sau*3AI
Xh	*Xho*I

The convention employed in the Figures is to show in parentheses a restriction enzyme site which was used for construction of the DNA sequence but was destroyed upon ligation of the construct.

Detailed Description of the Invention

In accordance with the present invention, a transformation procedure for the introduction of recombinant DNA material into host cells of the species *Pichia pastoris* is provided.

Further in accordance with the present invention, novel yeast strains of the species *Pichia pastoris* are provided which are useful as hosts for the introduction of recombinant DNA material.

Development of *Pichia pastoris* Transformation System

The transformation of *Pichia pastoris* has not been previously described. The experimental procedures for transformation of *Pichia pastoris* are presented in greater detail below (Example III). In order to develop a transformation system for *P. pastoris*, the auxotrophic mutant GS115 (NRRL Y-15851) was isolated and determined to be defective in the histidine pathway in that the strain has no detectable histidinol dehydrogenase activity. (See assay procedure in Example II).

Those of skill in the art recognize that mutation frequencies can be increased in a variety of ways, such as, for example, by subjecting exponentially growing cells to the action of a variety of mutagenic agents, such as, for example, N-methyl-N'-nitro-N-nitrosoguanidine, ethyl methanesulfonate, ultraviolet irradiation and the like. Isolation and identification of mutant strains defective in a specific metabolic pathway can be accomplished by determining the nutrient or nutrients required by the strain for growth as detailed, for example, in Example I. The specific gene and gene product in which a mutant strain is defective can then be determined by identifying the enzymatic activity which is absent, as detailed, for example, in Example II.

Yeast strains of the species *Pichia pastoris*, and especially mutant *Pichia pastoris* strains of the invention can be transformed by enzymatic digestion of the cell walls to give spheroplasts; the spheroplasts are then mixed with the transforming DNA and incubated in the presence of calcium ions and polyethylene glycol, then regenerated in selective growth medium. The transforming DNA includes the functional gene in which the host strain is defective, thus only transformed cells survive on the selective growth medium

employed.

To prepare *Pichia pastoris* spheroplasts, the cells are first contacted with a sulfhydryl group reducing agent, such as, for example, dithiothreitol or ß-mercaptoethanol. An example of a specific solution containing a sulfhydryl group reducing agent is the dithiothreitol in SED buffer described in the Examples. Enzymatic digestion of the cell walls can then be accomplished by contacting the strain to be transformed with any of the many cell wall degrading reagents known to those of skill in the art, such as for example Zymolyase (Miles Laboratories), Glusulase (Endo Laboratories), and the like. Although a wide variety of temperatures, contact times and dosage levels are operable, generally, when using, for example, zymolyase 60,000 (60,000 units/g) about 10 up to about 100 µg of cell wall degrading reagent per 10 mL of cell suspension are employed for spheroplast formation. Preferably about 40-50 µg of Zymolyase 60,000 per 10 mL of cell suspension is employed. Temperature is generally maintained at about 25° C or above, but less than about 35° C. Preferably, temperature is maintained at about 30° C. Contact time is generally at least about 15 minutes and usually no greater than about 60 minutes. While many buffered media are suitable, it is essential that cells to be converted to spheroplasts be suspended in a buffer which is iso-osmotic with the cells, such as, for example, SCE buffer (sorbitol/citrate/EDTA; see Examples for recipe).

The spheroplasts can be transformed by contact with virtually any amount of recombinant DNA material. Generally, at least about 0.01 µg of transforming DNA per 100 µL of spheroplast containing solution (containing between about 1-3x10^7 spheroplasts per 100 µL) are employed. Where only small amounts of recombinant DNA material are available, sonicated *E. coli* DNA can be used to supplement the amount of available DNA, thereby improving transformation frequencies by minimizing the handling losses of recombinant DNA material during experimental manipulation.

Transformed spheroplasts are then treated under cell wall regenerating conditions. Cell wall regenerating conditions comprise adding a sample containing transformed spheroplasts to melted regeneration agar maintained at about 40-60° C. A typical regeneration agar provides a balanced osmotic media and comprises:

sorbitol	about 1 M
dextrose	about 0.1 M
yeast nitrogen base	about 7 g/L
Bacto-agar	about 3%

The transformed spheroplasts in melted regeneration agar are poured over a bottom layer of regeneration agar and then incubated at about 25-35° C for about 3-10 days.

Pichia pastoris NRRL Y-15851 (GS115) has been transformed with a number of plasmids. Several of these plasmids are novel and have therefore been made available to the public by deposition with the Northern Regional Research Center in Peoria, Illinois. Plasmids and their accession numbers are tabulated below (all plasmids have been deposited in an *E. coli* host).

Plasmid	Inventor strain designation	NRRL accession number
pYA2	LE392-pYA2	B-15874
pYJ30	LE392-pYJ30	B-15890
pYJ32	LE392-pYJ32	B-15891
pSAOH5	MC1061-pSAOH5	B-15862

Also used to transform GS115 was plasmid pYA4, which is derived from the *S. cerevisiae* - *E. coli* shuttle vector YEp13 (available from ATCC #37115; see Figure 2). Thus, plasmid pYA4 is YEp13 plus a 6.0 kbp *Sau*3A partial digestion fragment of *Pichia pastoris* chromosomal DNA which includes the HIS4 gene (see Figure 3) ligated into the unique *Bam*HI site of YEp13.

Plasmid pYA2 (see Figure 1) contains pBR325 DNA sequences and a 9.3 kbp *S. cerevisiae Pst*I fragment which includes the *S. cerevisiae* HIS4 gene. It was surprisingly found that the *S. cerevisiae* HIS4

gene in plasmid pYA2 functioned in *Pichia pastoris*. An additional surprising observation was the fact that pYA2, which transforms *S. cerevisiae* at low frequency by integrative recombination, transformed *Pichia pastoris* at high frequency and was maintained as an extrachromosomal element in NRRL Y-15851 over a number of generations of growth.

Plasmid pYJ30, shown in Figure 4, has pBR322 DNA sequences, a 2.7 kbp *Bg1*II fragment of *Pichia pastoris* chromosomal DNA which has the *Pichia pastoris* HIS4 gene and a 164 bp *Taq*I fragment of *Pichia pastoris* chromosomal DNA which has autonomous replication sequence activity (PARS1). This plasmid has also been used to transform NRRL Y-15851 (GS115), and transformation occurs at high frequency. This plasmid is useful for introducing recombinant DNA material into a *Pichia pastoris* host. For example, plasmid pSAOH5 (see Figure 6) is derived from this plasmid by insertion of the *E. coli* LacZ gene and the alcohol oxidase regulatory region at the unique R_1 site of pYJ30. Plasmid pSAOH5 is shown in Example IV below to be capable of producing a polypeptide product not native to the host cell in *Pichia pastoris*.

Plasmid pYJ32, shown in Figure 5, is similar to pYJ30, except the autonomous replication activity is provided by PARS2, a 385 bp *Taq*I fragment of *Pichia pastoris* chromosomal DNA. This plasmid is also capable of transforming *Pichia pastoris* NRRL Y-15851 at high frequencies.

The transformation of yeast strains of the species *Pichia pastoris,* as demonstrated herein, makes possible the introduction of recombinant DNA material into yeast hosts. As further detailed in the examples which follow, transformed yeast strains of the species *Pichia pastoris* are useful, for example, for the production of polypeptide products by a yeast host.

EXAMPLES

The buffers and solutions employed in the following examples have the compositions given below:

1M Tris buffer	121.1 g Tris base in 800 mL of H_2O; adjust pH to the desired value by adding concentrated (35%) aqueous HC1; allow solution to cool to room temperature before final pH adjustment; dilute to a final volume of 1L.
TE buffer	1.0 mM EDTA in 0.01 M (pH 7.4) Tris buffer
YPD Medium	1% Bacto-yeast extract 2% Bacto-peptone 2% Dextrose
SD Medium	6.75 g yeast nitrogen base without amino acids (DIFCO) 2% Dextrose in 1 L of water
SED	1 M Sorbitol 25 mM EDTA 50 mM DTT
SCE Buffer	9.1 g Sorbitol 1.47 g Sodium citrate 0.168 g EDTA 50 mL H_2O --pH to 5.8 with HCl
CaS	1 M Sorbitol 10 mM $CaCl_2$ --filter sterilize
PEG Solution	20% polyethylene glycol-3350 10mM $CaCl_2$ 10mM Tris-HCl (pH 7.4) --filter sterilize
SOS	1 M Sorbitol 0.3x YPD medium 10 mM $CaCl_2$
MM (minimal medium)	

0.875 g	KH_2PO_4
0.125 g	K_2HPO_4
1.0 g	$(NH_4)_2SO_4$
0.5 g	$MgSO_4 \cdot 7H_2O$
0.1 g	NaCl
0.05 mg	$FeCl_3 \cdot 6H_2O$
0.07 mg	$ZnSO_4 \cdot 7H_2O$
0.01 mg	H_3BO_3
0.01 mg	$CuSO_4 \cdot 5H_2O$
0.01 mg	KI
0.1 g	$CaCl_2 \cdot 2H_2O$

--per liter of sterile H_2O

MM "minus"	MM formulation without $(NH_4)_2SO_4$
Citrate buffer	9.79 g sodium citrate 3.2 g citric acid ---dilute to 500 mL with H_2O ---adjust to pH 5.5 with 1 N NaOH
Nystatin solution	4.4 mg nystatin (5680 Units/mg) 1 mL dimethyl formamide ---dilute to 10 mL with water
E Buffer	50 mM Tris-HCl(pH 7.4) 0.01 mM histidinol 50 mM $MgSO_4$ 1 mM DTT
Vitamin Mix	

p-aminobenzoic acid	50 mg/100mL
p-hydroxybenzoic acid	50
riboflavin	25
pantothenate	50
B_{12}	1
folic acid	50
pyridoxine	50
biotin	5
thiamine	10
nicotinic acid	50
inositol	2000

The following abbreviations are used throughout the example, with the following meaning:

NTG	N-methyl-N'-nitro-N-nitrosoguanidine
DTT	dithiothreitol
NAD	nicotinamide adenine dinucleotide
SDS	sodium dodecyl sulfate
ala	alanine

arg	arginine
asn	asparagine
asp	aspartic acid
cys	cysteine
glu	glutamic acid
gln	glutamine
gly	glycine
his	histidine
ile	isoleucine
leu	leucine
lys	lysine
met	methionine
phe	phenylalanine
pro	proline
ser	serine
thr	threonine
trp	trytophan
tyr	tyrosine
val	valine

EXAMPLE I

Isolation of Auxotrophic Mutants

A. *Pichiapastoris* Mutagenesis

Culture of a selected yeast strain, such as for example, *Pichia pastoris* NRRL Y-11430, was inoculated into 100 mL of YPD broth and incubated at 30° C on a shaker for about 12-20 hrs. About 40 mL of the resulting culture were spun down at about 2,000 g for 5 minutes. The cells were then washed twice with 40 mL aliquots of sterile 0.1 M citrate buffer (pH 5.5). Washed cells were resuspended in 36 mL of sterile citrate buffer, then treated with 4 mL of NTG solution containing 5 mg of NTG per mL---thus giving a final NTG concentraton of 500 µg/mL. Cells in the presence of NTG were allowed to stand for about 30 minutes at room temperature without agitation.

NTG was then removed by washing the cells twice with 40 mL aliquots of sterile deionized water. Sufficient YPD medium was used to resuspend washed cells, which were then transferred to a flask and total volume brought up to 100 mL with additional YPD. These mutagenized cells were then incubated at 30° C on a shaker for about 48 hours.

After incubation, about 40 mL of the yeast containing solution were spun down at 2,000 g for 5 minutes. The cell pellet was washed twice with 40 mL aliquots of sterile, deionized water, then suspended in 40 mL of MM "minus" media plus 1% glucose carbon source and 5 µg biotin and incubated at 30° C on a shaker for 12-20 hours.

B. Nystatin enrichment

Five mL of the above culture grown on glucose was used to inoculate 100 mL of "restricted media". Restricted media comprises the MM formulation plus carbon source (typically 1% glucose), vitamin/amino acid supplementation as appropriate (such as the "vitamin mix" referred to above), except no supplementation is provided for the metabolite produced by the biosynthetic pathway in which a defect is sought. For example, where a leucine auxotroph is desired, no leucine supplementation will be provided. The inoculum in restricted media was incubated at 30° C in a shake flask and monitored periodically on a Klett-Summerson photoelectric colorimeter equipped with a 500-570 millimicron green filter. Incubation was continued until the scale reading (which is proportional to optical density) has increased 20-30% with respect to the original scale reading.

When the scale reading had increased as desired, the solution was treated with 1 mL of Nystatin solution, giving a Nystatin content of about 25 units/mL in the solution. The Nystatin-treated solution was incubated at 30° for 90 minutes without agitation, at which time 40 mL of the solution was spun down and the cells washed twice with 40 mL aliquots of deionized water. Washed cells were then diluted as appropriate in order to obtain about 100-150 colonies per plate. Colonies were plated on mutant growth

media which consisted of MM media, carbon source (typically 1% glucose), 5µg biotin and supplementation for any metabolite produced by the biosynthetic pathway in which the mutational defect is sought.

The colonies plated on mutant growth media were replica plated onto media formulation absent the metabolite supplementation. The original and replica plates were incubated at 30° for at least 48 hours. Those colonies that grew on the original plate (on mutant growth media) but not on the replica plates were selected for further characterization.

The auxotrophic mutants selected were transferred to metabolic pool plates and incubated at 30° C for at least 48 hours in order to determine in which pathway(s) mutational defects existed.

Pool plates were prepared by dissolving 10mg/mL of the L-isomer of each of 5 different amino acids, as follows:

	1	2	3	4	5
6	gly	asn	cys	met	gln
7	his	leu	ile	val	lys
8	phe	tyr	trp	thr	pro
9	glu	ser	ala	asp	arg

Thus, plate 1 contains 10 mg/mL each of glycine, histidine, phenylalanine and glutamic acid; plate 2 contains 10 mg/mL each of asparagine, leucine, tyrosine and serine, and so on. A tenth plate was prepared by dissolving 1 g of Casamino acids in 1 L of sterile water.

An aliquot of 250 µL of each of amino acid pools 1-10 was applied to plates containing minimal media plus 1% glucose, and the plates allowed to dry overnight.

The mutational defect of a given mutant can be determined by inspection of the growth pattern on the various pool plates. Thus GS115, a mutant defective in the histidine pathway, grew only on plates 1, 7 and 10, but not on the other pool plates which do not provide histidine supplementations. Similarly, GS190, a mutant defective in the arginine pathway, grew only on pool plates 5, 9 and 10, but did not grow on the other pool plates which did not have arginine supplementation.

EXAMPLE II

Identification of *Pichia pastoris* Mutants Defective in Histidinol Dehydrogenase Activity

A. Plate Test

Initial screening of histidine requiring mutants prepared as described in Example I was carried out to identify mutants defective at the *his*4C locus (i.e., lacking histidinol dehydrogenase activity). A master plate of histidine auxotrophic mutants was prepared with MM media, 1% glucose, vitamin mix (1 mL per L of media) and 0.2% Casamino acids. The master plate was incubated at 30° C for at least 48 hours, then four replica plates were prepared from the master plate:

(1) MM "minus" + 5µg biotin + 1% glucose + 0.2% histidinol
(2) MM media + 5µg biotin + 1% glucose + 0.0002 % histidinol
(3) MM "minus" + 5µg biotin + 1% glucose + 0.2% histidine
(4) MM media + 5µg biotin + 1% glucose + 0.002% histidine

These 4 plates were incubated at 30° C for at least 48 hours. Those colonies which grew on plates (3) and (4), but did not grow on plates (1) or (2) were selected for further analysis.

B. Enzymatic Analysis

The first step in the histidinol dehydrogenase assay procedure was to grow a 200 mL culture of a strain in YPD medium with shaking at 30° C to an OD_{600} of 1.0. The culture was then centrifuged at 2000 g for 5 minutes and the cells were resuspended in 200 mL of SD medium and incubated with shaking at 30° C. After 6-12 hours the culture was harvested by centrifugation and the cell pellet stored at -20° C.

The next step was to prepare a cell extract from the culture. Approximately 1 gram (wet weight) of cells was washed 2 times in 10 mL of cold H_2O (4° C) and resuspended in 0.83 mL of cold E buffer. To rupture

the cells, the sample was passed through an Aminco French pressure cell which had a 0.374 inch diameter piston using an Aminco French press at 20,000 PSI. The pressure cell was held on ice until use and the procedure was performed in a cold room (4° C). To monitor cell breakage, a 10 µL sample was added to 10 mL of H_2O and its OD_{600} determined and compared to an identically prepared control sample which had not been passed through the pressure cell. If the optical density of the treated sample was greater than 50% of the control, the sample was subjected to the disruption procedure a second time. The extract was then centrifuged in a Beckman SW50.1 rotor at 35,000 rpm and 4° C for 30 minutes to remove cell debris. The supernatant was removed, mixed with an equal volume of 4° C glycerol and stored at -20° C.

The concentration of total protein in an extract was estimated using the Bio-Rad Laboratories protein assay method. For this the Bio-Rad Dye Reagent Concentrate was diluted with four volumes of deionised H_2O and filtered through Whatman 3MM paper. A standard concentration curve was then prepared by adding 3, 10, 30, and 100 µg of bovine serum albumin (BSA) in 100 µL E buffer with 50% glycerol to a set of 13x100 mm glass tubes, each of which contained 2.5 mL of the dye reagent. The samples were mixed and held at room temperature for 5 minutes and their optical densities at 595 nm determined. For analyses of the extract, 3, 10, and 30 µL samples were brought to 100 µL with a solution containing E buffer and 50% glycerol and assayed for protein content as described above. A protein concentration value for each extract was then interpolated using the BSA concentration curve.

The final step in the histidinol dehydrogenase activity assay was to measure histidinol dehydrogenase activity in an extract by measuring spectrophotometerically the reduction of NAD which occurs in the presence of histidinol. For each extract to be assayed, a reaction mixture which contained 3 mL of H_2O, 0.5 mL of 0.5 M glycine (pH 9.4), 0.5 mL of 5 mM $MnCl_2$ and 0.5 mL of 0.1 M NAD was prepared on ice. A 2.25 mL aliquot of this mix was added to 2 13x100 mm glass tubes which were on ice. A sample which contained between 50 to 500 µg of protein was added to each of the tubes and the tubes were incubated at 25° C. After 5 minutes the reaction was started by the addition of 0.25 mL of 0.15 M histidinol to one tube and 0.25 mL of H_2O to the other. The optical density of each reaction tube at 340 nm was determined at times of 0.0, 0.5, 1.0 and 2.0 hours. As controls, extracts prepared from *Pichia pastoris* NRRL Y-11430 and *Saccharomyces cerevisiae* 5799-4D (NRRL Y-15859) were assayed in parallel. The net OD_{340} value for each time point was determined by subtracting the value obtained from the sample incubated without histidinol from the value obtained from the sample incubated with histidinol.

While *Pichia pastoris* NRRL Y-11430, a wild type strain requiring no amino acid supplementation, gave an OD_{340} of about 0.25, 0.38 and 0.75 at 0.5, 1.0 and 2.0 hours, respectively, the control *his*4C mutant (*S. cerevisiae* NRRL Y-15859) gave an OD_{340} of essentially zero at all time points. One such *Pichia pastoris* mutant, designated GS115 and deposited with the Northern Regional Research Center having the accession number NRRL Y-15851, similarly gave an OD_{340} of essentially zero at all time points. Consistent with the mutant genotype nomenclature employed for *S. cerevisiae*, GS115 has been designated as a *his*4C mutant strain.

EXAMPLE III

Pichiapastoris Transformation Procedure

A. Cell Growth

1. Inoculate a colony of *Pichia pastoris* GS115 (NRRL Y-15851) into about 10 mL of YPD medium and shake culture at 30° C for 12-20 hrs.
2. After about 12-20 hrs., dilute cells to an OD_{600} of about 0.01-0.1 and maintain cells in log growth phase in YPD medium at 30° C for about 6-8 hrs.
3. After about 6-8 hrs, inoculate 100 mL of YPD medium with 0.5 mL of the seed culture at an OD_{600} of about 0.1 (or equivalent amount). Shake at 30° C for about 12-20 hrs.
4. Harvest culture when OD_{600} is about 0.2-0.3 (after approximately 16-20 hrs) by centrifugation at 1500 g for 5 minutes.

B. Preparation of Spheroplasts

1. Wash cells once in 10 mL of sterile water. (All centrifugations for steps 1-5 are at 1500 g for 5 minutes.)
2. Wash cells once in 10 mL of freshly prepared SED.
3. Wash cells twice in 10 mL of sterile 1 M Sorbitol.

4. Resuspend cells in 10 mL SCE buffer.
5. Add 5-10 µL of 4 mg/mL Zymolyase 60,000 (Miles Laboratories). Incubate cells at 30° C for about 30-60 minutes.

Since the preparation of spheroplasts is a critical step in the transformation procedure, one should monitor spheroplast formation as follows: add 100 µL aliquots of cells to 900 µL of 5% SDS and 900 µL of 1 M Sorbitol before or just after the addition of zymolyase and at various times during the incubation period. Stop the incubation at the point where cells lyse in SDS but not in sorbitol (usually between 30 and 60 minutes of incubation).

6. Wash spheroplasts twice in 10 mL of sterile 1 M Sorbitol by centrifugation at 1000 g for 5-10 minutes. (The time and speed for centrifugation may vary; centrifuge enough to pellet spheroplasts but not so much that they rupture from the force.)
7. Wash cells once in 10 mL of sterile CaS.
8. Resuspend cells in total of 0.6 mL of CaS.

C. Transformation

1. Add DNA samples (up to 20 µL volume) to 12 X 75 mm sterile polypropylene tubes. (DNA should be in water or TE buffer; for maximum transformation frequencies with small amounts of DNA, it is advisable to add about 1 µL of 5 mg/mL sonicated *E. coli* DNA to each sample.)
2. Add 100 µL of spheroplasts to each DNA sample and incubate at room temperature for about 20 minutes.
3. Add 1 mL of PEG solution to each sample and incubate at room temperature for about 15 minutes.
4. Centrifuge samples at 1000 g for 5-10 minutes and decant PEG solution.
5. Resuspend samples in 150 µL of SOS and incubate for 30 minutes at room temperature.
6. Add 850 µL of sterile 1 M Sorbitol and plate aliquots of samples as described below.

D. Regeneration of Spheroplasts

1. Recipe for Regeneration Agar Medium:
   a. Agar-Sorbitol- 9 g Bacto-agar, 54.6 g Sorbitol, 240 mL H_2O, autoclave.
   b. 10X Glucose- 20 g Dextrose, 100 mL H_2O, autoclave.
   c. 10X SC- 6.75 g Yeast Nitrogen Base without amino acids, 100 mL H_2O, autoclave. (Add any desired amino acid or nucleic acid up to a concentration of 200 µg/mL before or after autoclaving.)
   d. Add 30 mL of 10X Glucose and 30 mL of 10X SC to the melted Agar-Sorbitol solution to give a total of 300 mL. Add 0.6 mL of 0.2 mg/mL biotin and any other desired amino acid or nucleic acid to a concentration of 20 µg/mL. Hold melted Regeneration Agar at 55-60° C.
2. Plating of Transformation Samples:

Pour bottom agar layer of 10 mL Regeneration Agar per plate at least 30 minutes before transformation samples are ready. Distribute 10 mL aliquots of Regeneration Agar to tubes in a 45-50° C bath during the period that transformation samples are in SOS. Add aliquots of transformation samples described above to tubes with Regeneration Agar and pour onto bottom agar layer of plates. Add a quantity of each sample to 10 mL aliquots of melted Regeneration Agar held at 45-50° C and pour each onto plates containing a solid 10 mL bottom agar layer of Regeneration Agar.

3. Determination of Quality of Spheroplast Preparation:

Remove 10 µL of one sample and dilute 100 times by addition to 990 µL of 1 M Sorbitol. Remove 10 µL of the 100 fold dilution and dilute an additional 100 times by addition to a second 990 µL aliquot of 1 M Sorbitol. Spread plate 100 µL of both dilutions on YPD agar medium to determine the concentration of unspheroplasted whole cells remaining in the preparation. Add 100 µL of each dilution to 10 mL of Regeneration Agar supplemented with 40 µg/mL histidine to determine total regeneratable spheroplasts. Good values for a transformation experiment are 1-3 X 10^7 total regeneratable spheroplasts/mL and about 1 X 10^3 whole cells/mL.

4. Incubate plates at 30° C for 3-5 days.

EXAMPLE IV

Production of β-Galactosidase in *Pichia pastoris*

The production of β-galactosidase in transformed *Pichia pastoris* demonstrates the ability of yeast of

the species *Pichia pastoris* to be employed as a host/vector system for the production of polypeptide products. *Pichia pastoris* GS115 (NRRL Y-15851) was transformed with plasmid pSAOH5 (see Figure 6) and grown up in minimal medium containing 0.5 µg/mL of biotin and 0.1% glucose at 30°C until they reached stationary phase. The cells were then shifted to minimal medium containing 0.5 µg/mL of biotin and 0.5% methanol and grown for about 3-5 generations at 30°C. After this initial growth on methanol, cells were shifted to fresh minimal media containing 0.5 µg/mL biotin and 0.2% methanol as carbon source. The cells were incubated at 30°C for about 80 hours, with samples periodically drawn to determine alcohol oxidase and β-galactosidase levels.

The first sample drawn, immediately after the cells were shifted to the growth medium, analyzed for over 500 units of alcohol oxidase and over 1100 units of β-galactosidase. Assay procedures employed are detailed in the appendix.

These results demonstrate the utility of of *Pichia pastoris* as a host/vector system for the production of gene products in yeast. The plasmid employed to transform the host, plasmid pSAOH5, is a *Pichia* plasmid which codes for the production of β-galactosidase under the control of a methanol responsive regulatory region. The transformed strain used for this demonstration has been deposited with the Northern Regional Research Center and is available to the public under the accession number NRRL Y-15853.

APPENDIX

Alcohol Oxidase Assay

The alcohol oxidase activity for reaction with methanol was determined by the following assay procedure (dye-peroxidase method). A dye-buffer mixture was prepared by mixing 0.1 mL of an o-dianisidine solution (1 weight % o-dianisidine in water) with 12 mL of aerated 0.1 M sodium phosphate buffer (pH 7.5). The assay mixture was prepared with 2.5 mL of the dye-buffer mixture, 50 µL of methanol, 10 µL of a peroxidase solution (1 mg of horse-radish peroxidase-Sigma, Type II), and 25 µL of the alcohol oxidase solution. The assay mixture was maintained at 25°C in a 4x1x1 cm cuvette and the increase in absorbance by the dye at 460 nm was recorded for 2 to 4 minutes. The enzyme activity was calculated by

$$\text{Activity (}\mu\text{ mole/min/mL or Enzyme Units/mL)} = \frac{\Delta A}{\text{min}} \times 11.5$$

wherein 11.5 is a factor based on a standard curve prepared with known aliquots of H_2O_2 and ΔA is the change in absorbance during the experimental interval.

β-Galactosidase Assay

β-Galactosidase was determined as follows:

A. Solutions required:

Z-buffer:

		Final concentration
$Na_2HPO_4 \cdot 7H_2O$	16.1 g	0.06 M
NaH_2PO_4	5.5 g	0.04 M
KCl	0.75 g	0.01 M
$MgSO_4 \cdot 7H_2O$	0.246 g	0.001 M
2-mercaptoethanol	2.7 mL	0.05 M

fill up to 1L; pH should be 7

O-Nitrophenyl-β-D-galactoside (ONPG):

Dissolve 400 mg ONPG (Sigma N-1127) in 100 mL of distilled water to make a 4 mg/mL ONPG solution.

B. Assay Procedure:

1. Withdraw an aliquot from the culture medium (0.1-0.5 OD_{600} of yeast cells), centrifuge and wash cell pellet with water.
2. Add 1 mL of Z buffer to the cell pellet, 30 µL of $CHCl_3$ and 30 µL of 0.1% SDS, vortex, incubate 5 minutes at 30° C.
3. Start reaction by adding 0.2 mL of ONPG (4 mg/mL), vortex.
4. Stop reaction by adding 0.5 mL of a 1 M Na_2CO_3 solution at appropriate time points $A_{420}<1$).
5. Read absorbance of supernatant at 420 nm.

C. Calculation of β-galactosidase Units:

1 U = 1 nmole of orthonitrophenol (ONP) formed per minute at 30° C and a pH 7.

1 nmole of ONP has an absorbance at 420 nm (A_{420}) of 0.0045 with a 1 cm pathlength; thus, an absorbance of 1 at 420 nm represents 222 nmole ONP/mL, or 378 nmole/1.7 mL since the total volume of supernatant being analyzed is 1.7 mL. Hence, Units are calculated as follows:

$$U = \frac{A_{420}}{t(min)} \times 378$$

The following part of the description are preferred embodiments 1 to 30 presented in the format of claims.

Claims

1. A yeast cell of the species Pichia pastoris as a host capable of being transformed with recombinant DNA material; wherein said host is defective in histidine biosynthetic pathway.

2. The yeast cell of claim 1 characterized in that said histidine biosynthetic pathway is defective in histidinol dehydrogenase activity.

3. The yeast cell of claim 1 characterized in that said recombinant DNA material comprises a functional gene which complements the defect in the histidine biosynthetic pathway in which the host is defective.

4. Pichia pastoris NRRLY-15851 (GS115)

5. A process for transforming a host yeast strain of Pichia pastoris, said process comprising:
   (a) contacting the host yeast strain with a sulfhydryl group reducing agent;
   (b) contacting the product cells of step (a) with a cell wall degrading reagent under conditions suitable for the formation and maintenance of spheroplasts;
   (c) contacting the spheroplasts generated in step (b) with recombinant DNA material under conditions suitable for transformation; and
   (d) treating the product of step (c) under cell wall regenerating conditions.

6. The process of claim 5 wherein said sulfhydryl group reducing agent is dithiothreitol.

7. The process of claim 5 wherein said cell wall degrading reagent is Zymolyase.

8. The process of claim 5 wherein said conditions suitable for the formation of spheroplasts comprise:
   (i) 10-100 µg of cell wall degrading reagent per 10 mL of cell suspension; wherein said cell suspension is prepared by suspending exponentially growing cells in Sorbitol-Citrate-EDTA buffer, pH 5.8 (SCE buffer)

(ii) maintained at 25-35 ° C;
(iii) for 15-60 minutes.

9. The process of any claims 5 to 8 characterized in that said host yeast strain is defective in histidine biosynthetic pathway.

10. The process of claim 9 characterized in that said histidine biosynthetic pathway is defective at the gene encoding histidinol dehydrogenase.

11. The process of claim 10 wherein said host yeast strain is Pichia pastoris NRRL Y-15851 (GS115).

12. The process of claim 5 characterized in that said contacting conditions comprise:
(i) 2-10 volumes of $CaCl_2$-polyethylene glycol solution per volume of spheroplast-containing suspension;
(ii) maintained at 20-30 ° C;
(iii) for 5-30 minutes.

13. The process of claim 12 wherein said cell wall regenerating conditions comprise:
(i) adding transformed spheroplasts to regeneration agar, wherein said regeneration agar comprises:
about 1 M sorbitol,
about 0.1 M dextrose,
about 7 g/L yeast nitrogen base, and
about 3% agar;
(ii) maintained at 25-35 ° C;
(iii) for about 3-10 days.

14. The process of claim 9 characterized in that said recombinant DNA material comprises a functional gene which complements the defect in the histidine biosynthetic pathway in which the host yeast strain is defective.

15. The process of claim 14 wherein said recombinant DNA material comprises a histidinol dehydrogenase encoding gene.

16. The process of claim 14 wherein said recombinant DNA material is plasmid pYA2(NRRL B-1587).

17. The process of claim 14 wherein said recombinant DNA material is plasmid pYA4, which is YEp13 (ATCC 37115) plus a 6.0 kbp Sau3A partial digestion fragment of Pichia pastoris chromosomal DNA which includes the His4 gene ligated into the unique BamHI site of YEp13.

18. The process of claim 14 wherein said recombinant DNA material is plasmid pYJ30 (NRRL B-15890).

19. The process of claim 14 wherein said recombinant DNA material is plasmid pYJ32 (NRRL B-15891).

20. The process of claim 5 further comprising:
(e) growing the product cells of step (d) under selective growth conditions.

21. The process of claim 20 further comprising:
(e) growing the product cells of step (d) under selective growth conditions; wherein said selective growth conditions comprise growth on yeast minimal medium without added histidine.

Revendications

1. Cellule de levure de l'espèce Pichia pastoris comme hôte susceptible d'être transformé par du matériau d'ADN recombinant; où la voie de la biosynthèse de l'histidine de l'hôte est défectueuse.

2. Cellule de levure selon la revendication 1, caractérisée en ce que ladite voie de la biosynthèse de l'histidine est défectueuse au niveau de l'activité histidinol déshydrogénase.

3. Cellule de levure selon la revendication 1, caractérisée en ce que ledit matériau d'ADN recombinant comprend un gène fonctionnel qui compense le défaut dans la voie de la biosynthèse de l'histidine de l'hôte.

4. Pichia pastoris NRRLY-15851 (GS115).

5. Procédé pour transformer une souche de levures hôtes Pichia pastoris, ledit procédé comprenant de :
(a) mettre en contact la souche de levures hôtes avec un agent réducteur à groupes sulfhydryle;
(b) mettre en contact les cellules produites à l'étape (a) avec un réactif capable de dégrader les parois cellulaires dans des conditions permettant de former et de conserver des sphéroplastes;
(c) mettre en contact les sphéroplastes produits à l'étape (b) avec du matériau d'ADN recombinant dans des conditions permettant leur transformation; et
(d) traiter le produit de l'étape (c) dans des conditions de régénération des parois cellulaires.

6. Procédé selon la revendication 5, dans lequel ledit agent réducteur à groupes sulfhydryle est le dithiothréitol.

7. Procédé selon la revendication 5, dans lequel ledit agent capable de dégrader les parois cellulaires est la Zymolyase.

8. Procédé selon la revendication 5, dans lequel les conditions appropriées permettant de former des sphéroplastes comprennent :
(i) 10-100 µg de réactif capable de dégrader les parois cellulaires par 10 ml de suspension cellulaire; où ladite suspension cellulaire est préparée en mettant en suspension des cellules en croissance exponentielle dans un tampon sorbitol-citrate-EDTA pH 5,8 (tampon SCE);
(ii) maintenir à 25-35 °C;
(iii) pendant 15-60 minutes.

9. Procédé selon l'une quelconque des revendications de 5 à 8 caractérisé en ce que la souche de levures hôtes présente un défaut dans la voie de la biosynthèse de l'histidine.

10. Procédé selon la revendication 9, caractérisé en ce que ladite voie de biosynthèse de l'histidine présente un défaut dans le gène codant l'histidinol déshydrogénase.

11. Procédé selon la revendication 10, dans lequel ladite souche de levure hôtes est Pichia pastoris NRRL Y-15851 (G5115).

12. Procédé selon la revendication 5, caractérisé en ce que lesdites conditions de contact comprennent :
(i) 2-10 volumes de solution $CaCl_2$-polyéthylène glycol par volume de suspension des sphéroplastes;
(ii) maintenir à 20-30 °C;
(iii) pendant 5-30 minutes.

13. Procédé selon la revendication 12, dans lequel lesdites conditions de régénération des parois cellulaires comprennent de :
(i) ajouter les sphéroplastes transformés à de l'agar de régénération, lequel agar de régénération comprend :
environ 1 M de sorbitol
environ 0,1 M de dextrose
environ 7 g/l d'une base azotée pour levure, et
environ 3% d'agar;
(ii) maintenir à 25-35 °C;
(iii) pendant environ 3-10 jours.

14. Procédé selon la revendication 9, caractérisé en ce que ledit matériau d'ADN recombinant comprend un gène fonctionnel qui compense le défaut dans la voie de biosynthèse de l'histidine manque dans la souche de levures hôtes.

15. Procédé selon la revendication 14, dans lequel ledit matériau d'ADN recombinant comprend un gène codant l'histidinol déshydrogénase.

16. Procédé selon la revendication 14, dans lequel ledit matériau d'ADN recombinant est le plasmide pYA2 (NRRL B-15874.

17. Procédé selon la revendication 14, dans lequel ledit matériau d'ADN recombinant est le plasmide pYA4, qui est UN YEp13 (ATCC 37115) plus un fragment de digestion partielle d'ADN chromosomique Sau3A de 6,0 kilopaires de bases de Pichia pastoris, qui comprend le gène His4 ligaturé dans l'unique site BamHI de YEp13.

18. Procédé selon la revendication 14, dans lequel ledit matériau d'ADN recombinant est le plasmide pYJ30 (NRRL B-15890).

19. Procédé selon la revendication 14, dans lequel ledit matériau d'ADN recombinant est le plasmide pYJ32 (NRRL B-15891).

20. Procédé selon la revendication 5 comprenant en outre de :
(e) cultiver les cellules produites à l'étape (d) dans des conditions de croissance sélective.

21. Procédé selon la revendication 2, comprenant en outre de :
(e) cultiver les cellules produites à l'étape (d) dans des conditions de croissance sélectives; lesquelles conditions de croissance sélective comprennent une croissance sur un milieu minimum pour levures sans ajout d'histidine.

Patentansprüche

1. Hefezelle der Spezies Pichia pastoris als Wirt, der mit rekombinantem DNA-Material transformiert werden kann, wobei der Wirt einen Defekt im biosynthetischen Histidinstoffwechsel aufweist.

2. Hefezelle nach Anspruch 1, dadurch gekennzeichnet, dass der biosthetische Histidinstoffwechsel einen Defekt an Histidinol-dehydrogenase-Aktivität aufweist.

3. Hefezelle nach Anspruch 1, dadurch gekennzeichnet, dass das rekombinante DNA-Material ein funktionelles Gen umfasst, das den Defekt im biosynthetischen Histidinstoffwechsel, in dem der Wirt einen Defekt aufweist, komplementiert.

4. Pichia pastoris NRRL Y-15851 (GS115).

5. Verfahren zur Transformation eines Wirtshefestamms von Pichia pastoris, wobei das Verfahren folgendes umfasst:
(a) Kontaktieren des Wirtshefestamms mit einem Sulfhydrylgruppen-Reduktionsmittel;
(b) Kontaktieren der in Stufe (a) als; Produkt gebildeten Zellen mit einem zellwandabbauenden Mittel unter Bedingungen, die zur Bildung und Aufrechterhaltung von Sphäroplasten geeignet sind;
(c) Kontaktieren der in Stufe (b) gebildeten Sphäroplasten mit rekombinantem DNA-Material unter für die Transformation geeigneten Bedingungen; und
(d) Behandeln des Produkts von Stufe (c) unter zellwandregenerierenden Bedingungen.

6. Verfahren nach Anspruch 5, wobei es sich beim Sulfhydrylgruppen-Reduktionsmittel um Dithiothreit handelt.

7. Verfahren nach Anspruch 5, wobei es sich beim zellwandabbauenden Mittel um Zymolyase handelt.

8. Verfahren nach Anspruch 5, wobei die zur Bildung von Sphäroplasten geeigneten Bedingungen folgendes umfassen:
(i) 10 - 100 µg zellwandabbauendes Mittel pro 10 ml Zellsuspension; wobei die Zellsuspension durch Suspendieren von exponentiell wachsenden Zellen in Sorbit-Citrat-EDTA-Puffer vom pH-Wert 5,8 (SCE-Puffer) hergestellt worden ist;

(ii) Belassen bei 25 - 35 °C;
(iii) für 15 - 60 Minuten.

9. Verfahren nach einem der Ansprüche 5 bis 8, dadurch gekennzeichnet, dass der Wirtshefestamm einen Defekt im biosynthetischen Histidinstoffwechsel aufweist.

10. Verfahren nach Anspruch 9, dadurch gekennzeichnet, dass der biosynthetische Histidinstoffwechsel einen Defekt am für Histidinoldehydrogenase kodierenden Gen aufweist.

11. Verfahren nach Anspruch 10, wobei es sich beim Wirtshefestamm um Pichia pastoris NRRL Y-15851 (GS115) handelt.

12. Verfahren nach Anspruch 5, dadurch gekennzeichnet, dass die Kontaktbedingungen folgendes umfassen:
(i) 2 - 10 Volumenteile $CaCl_2$-Polyethylenglykollösung pro Volumenteil der Sphäroplasten enthaltenden Suspension;
(ii) Belassen bei 20 - 30 °C;
(iii) für 5 - 30 Minuten.

13. Verfahren nach Anspruch 12, wobei die zellwandregenerierenden Bedingungen folgendes umfassen:
(i) Zugabe von transformierten Sphäroplasten zu Regenerationsagar, wobei das Regenerationsagar folgendes enthält:
etwa 1 m Sorbit,
etwa 0,1 m Dextrose,
etwa 7 g/l Hefe-Stickstoffbase und
etwa 3 % Agar;
(ii) Belassen bei 25 - 35 °C;
(iii) für etwa 3 - 10 Tage.

14. Verfahren nach Anspruch 9, dadurch gekennzeichnet, dass das rekombinante DNA-Material ein funktionelles Gen umfasst, das den Defekt im biosynthetischen Histidinstoffwechsel, den der Wirtshefestamm aufweist, komplementiert.

15. Verfahren nach Anspruch 14, wobei das rekombinante DNA-Material ein für Histidinol-dehydrogenase kodierendes Gen umfasst.

16. Verfahren nach Anspruch 14, wobei es sich beim rekombinanten DNA-Material um das Plasmid pYA2 (NRRL B-15874) handelt.

17. Verfahren nach Anspruch 14, wobei das rekombinante DNA-Material das Plasmid pYA4 ist, wobei es sich um YEp13 (ATCC 37115) plus ein 6,0 kbp Sau3A-partielles Verdauungsfragment von Pichia pastoris-chromosomaler DNA, das das in die einzige BamHI-Stelle von YEp13 ligierte His4 einschliesst, handelt.

18. Verfahren nach Anspruch 14, wobei es sich bei dem rekombinanten DNA-Material um das Plasmid pYJ30 (NRRL B-15890) handelt.

19. Verfahren nach Anspruch 14, wobei es sich bei dem rekombinanten DNA-Material um das Plasmid pYJ32 (NRRL B-15891) handelt.

20. Verfahren nach Anspruch 5, zusätzlich umfassend:
(e) Züchtung der Produktzellen von Stufe (d) unter selektiven Wachstumsbedingungen.

21. Verfahren nach Anspruch 20, zusätzlich umfassend:
(e) Züchtung der Produktzellen von Stufe (d) unter selektiven Wachstumsbedingungen; wobei diese selektiven Wachstumsbedingungen die Züchtung auf Hefe-Minimalmedium ohne Histidinzusatz umfassen.

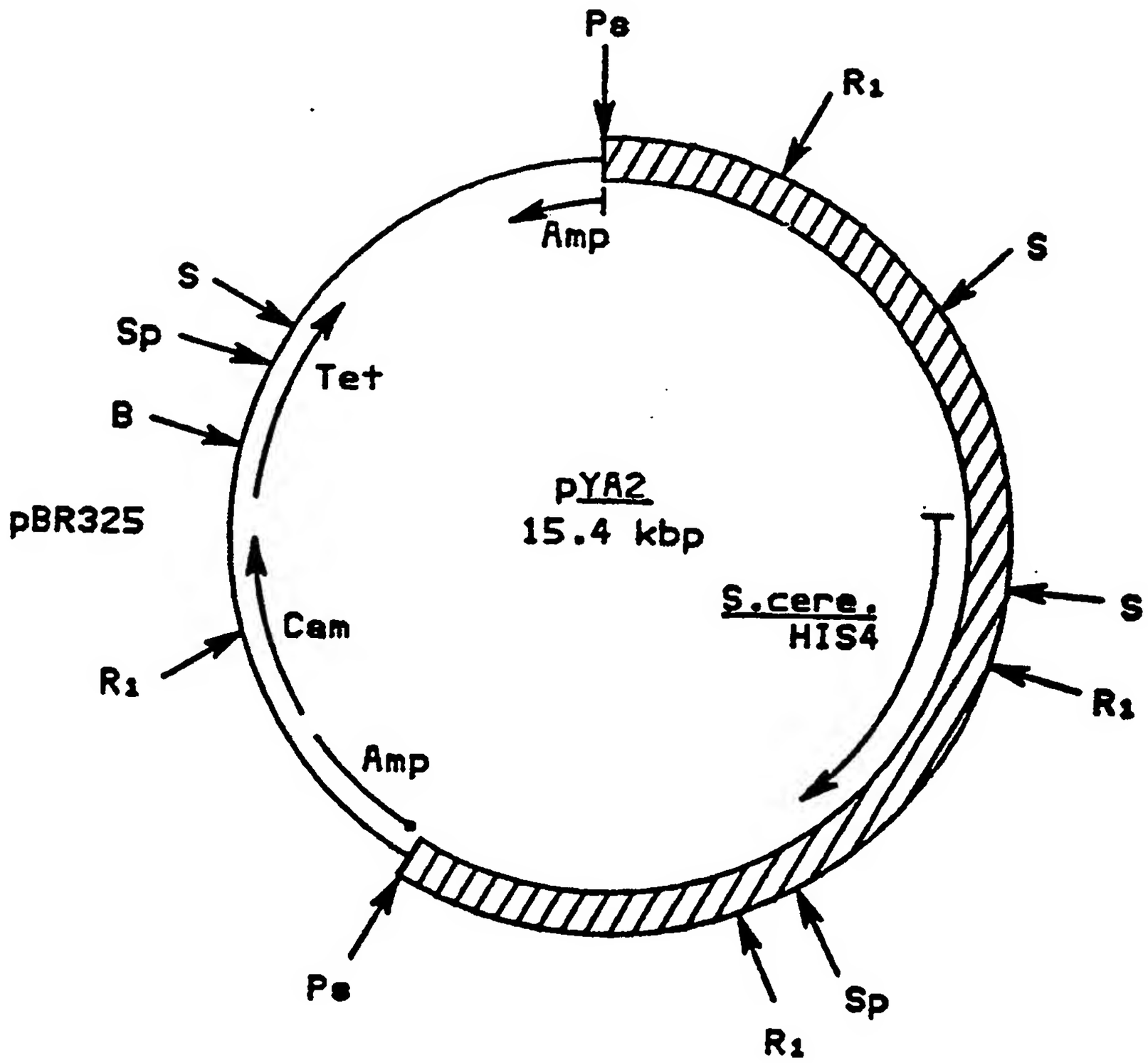
Pe
R1
Amp
S
S
Sp
B
Tet
pBR325
pYA2
15.4 kbp
S.cere.
HIS4
S
R1
Cam
R1
Amp
Pe
Sp
R1

FIG. 1

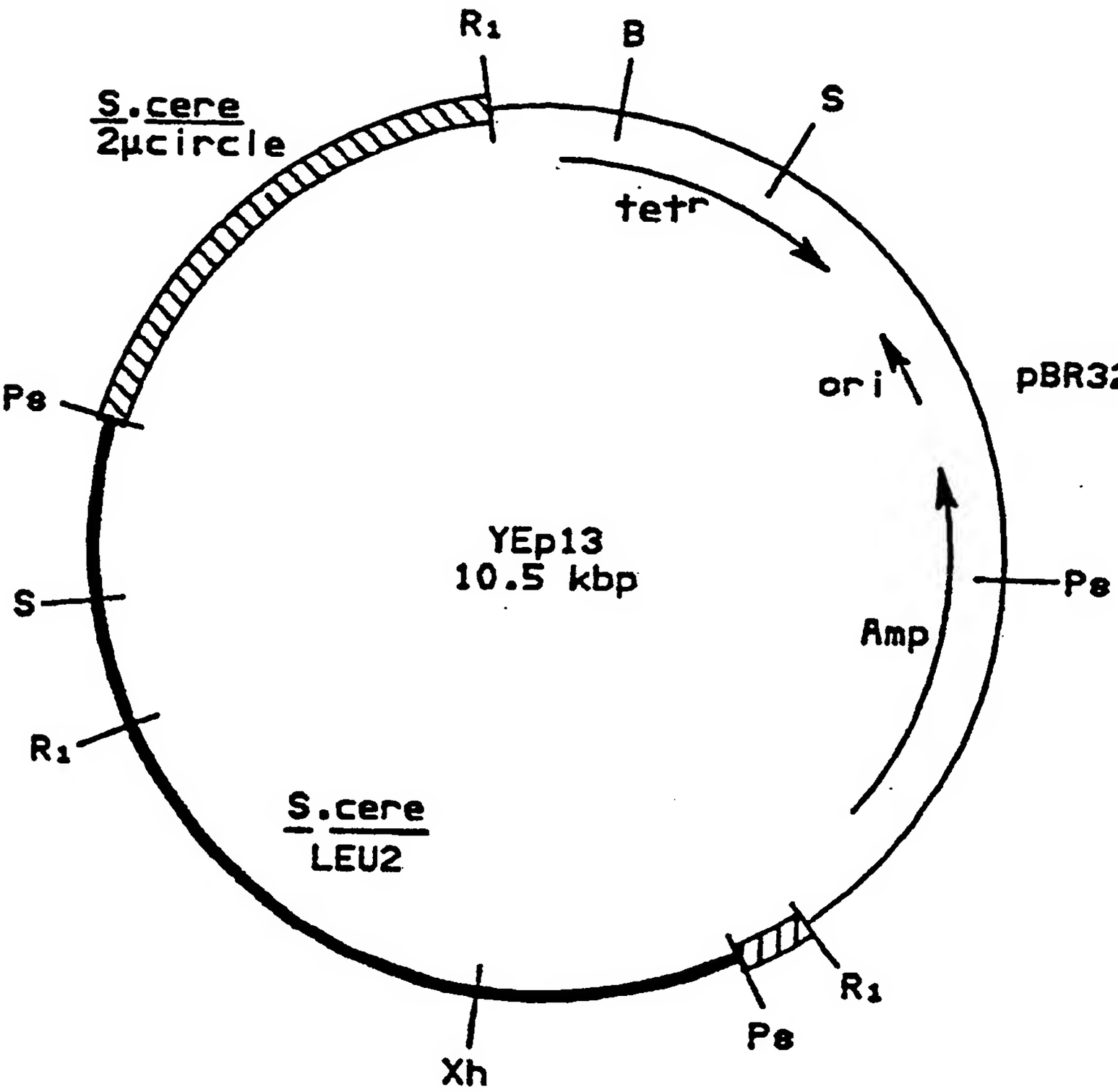
R1
B
S
S.cere
2µcircle
tetr
ori
pBR322
Pe
YEp13
10.5 kbp
Amp
Pe
S
R1
S.cere
LEU2
R1
Pe
Xh

FIG. 2

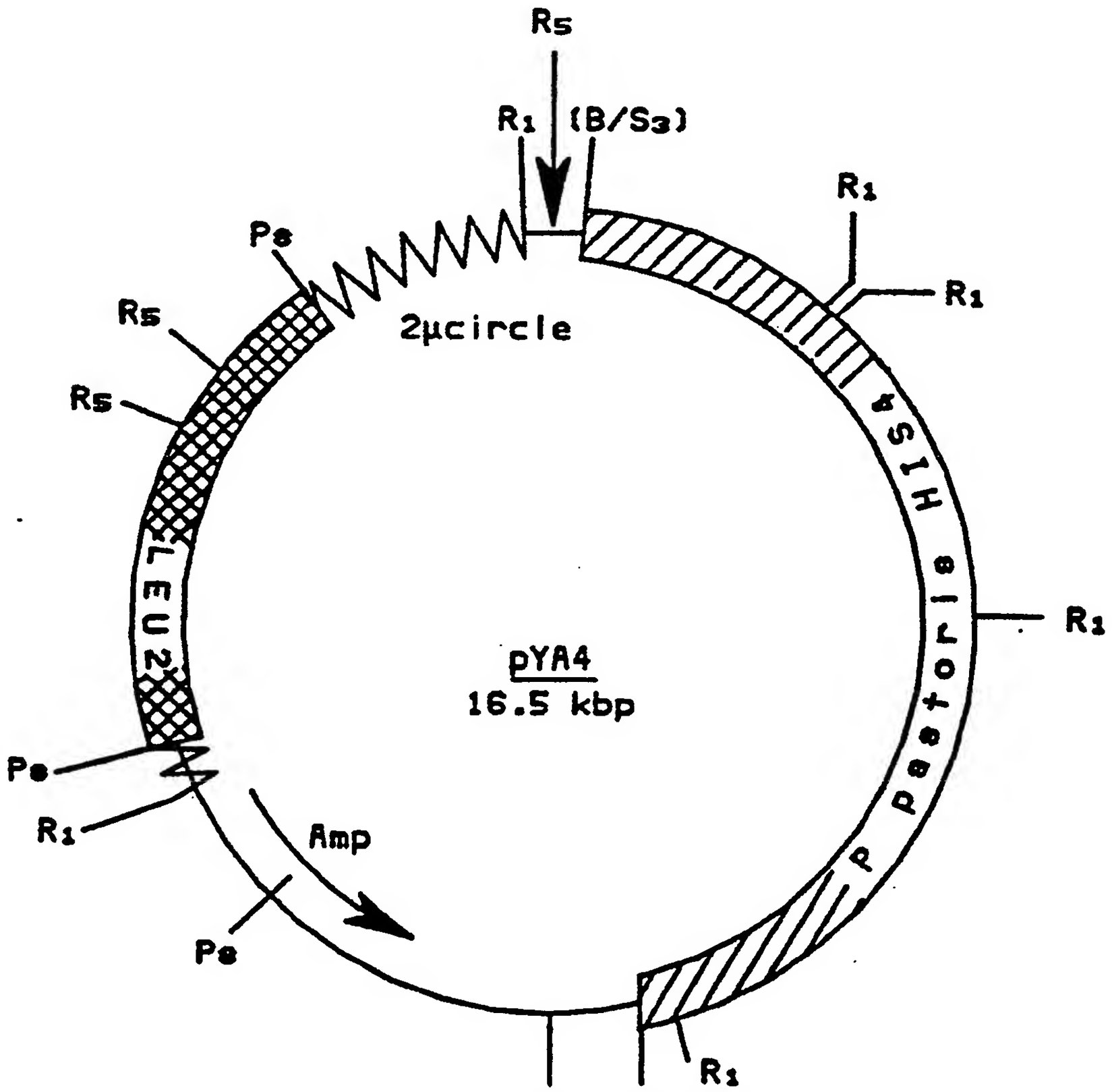
Rs
R1 (B/S3)
R1
Pe
R1
Rs
2µcircle
Rs
HIS4
LEU2
R1
pYA4
16.5 kbp
Pe
R1
Amp
Pe
R1
Sp (B/S3)

FIG. 3

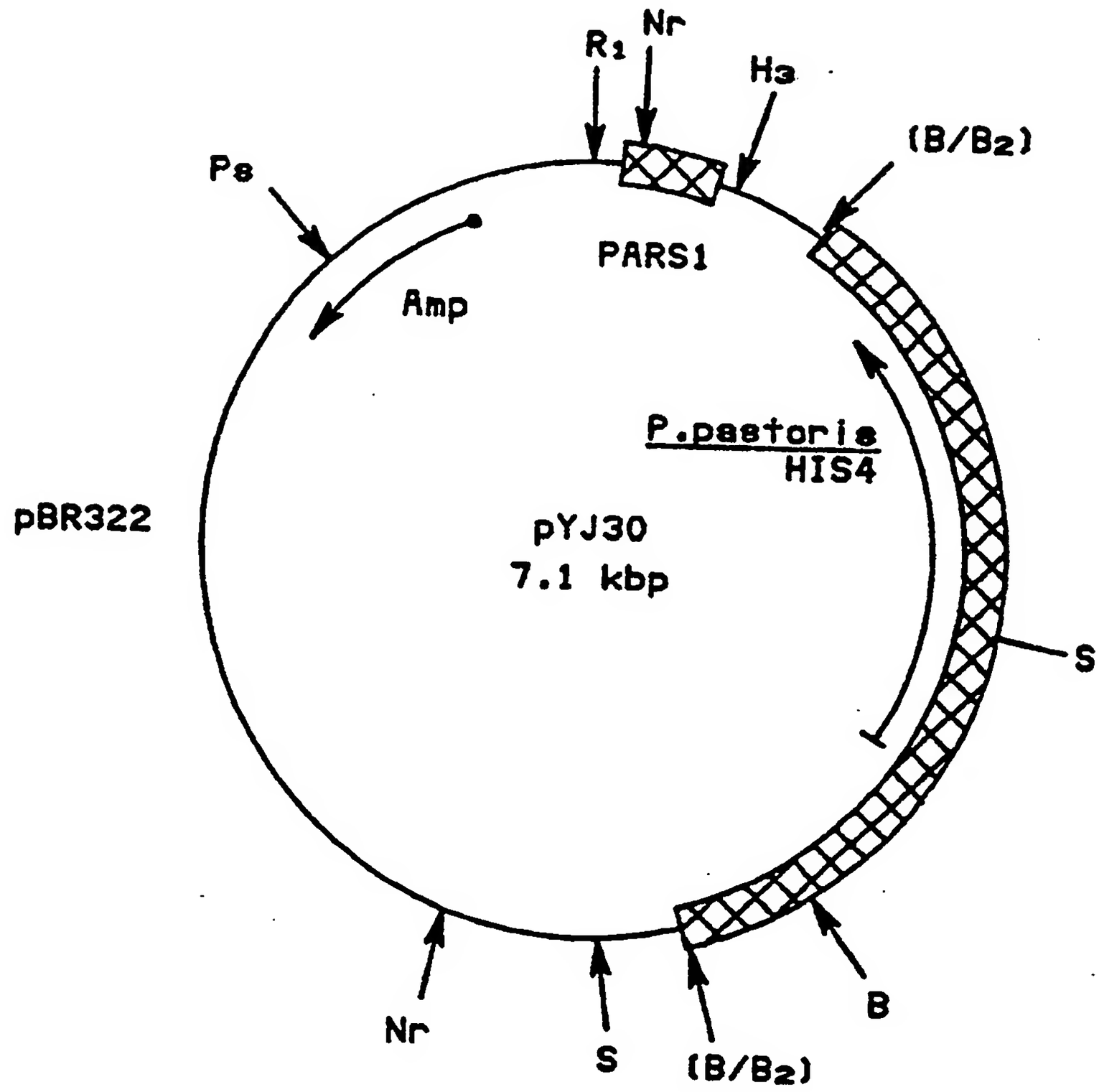
R1
Nr
H3
(B/B2)
Ps
PARS1
Amp
P.pastoris
HIS4
pBR322
pYJ30
7.1 kbp
S
Nr
B
S
(B/B2)

FIG. 4

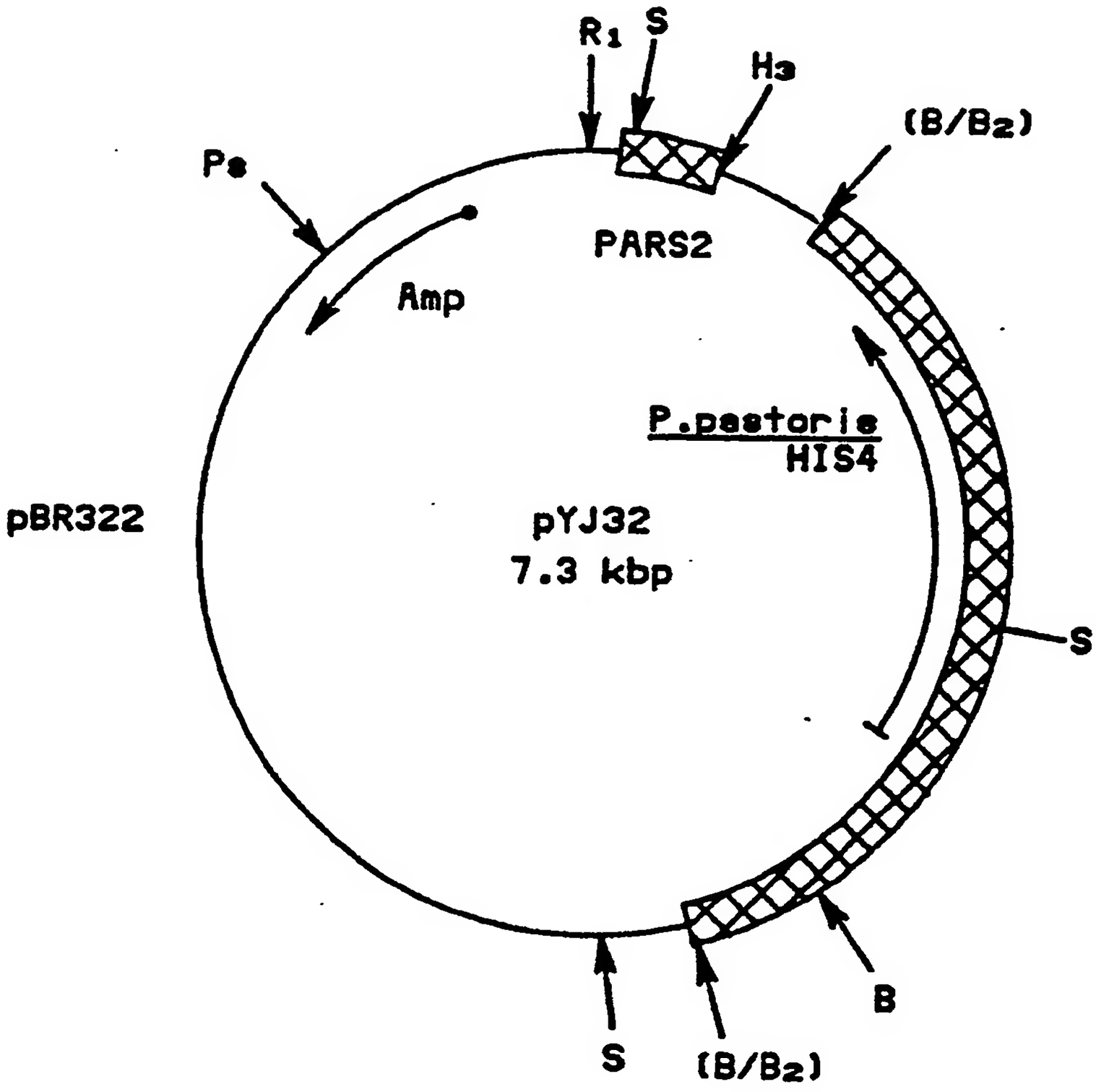
R1
S
H3
(B/B2)
P
PARS2
Amp
P.pastoris
HIS4
pBR322
pYJ32
7.3 kbp
S
B
S
(B/B2)

FIG. 5

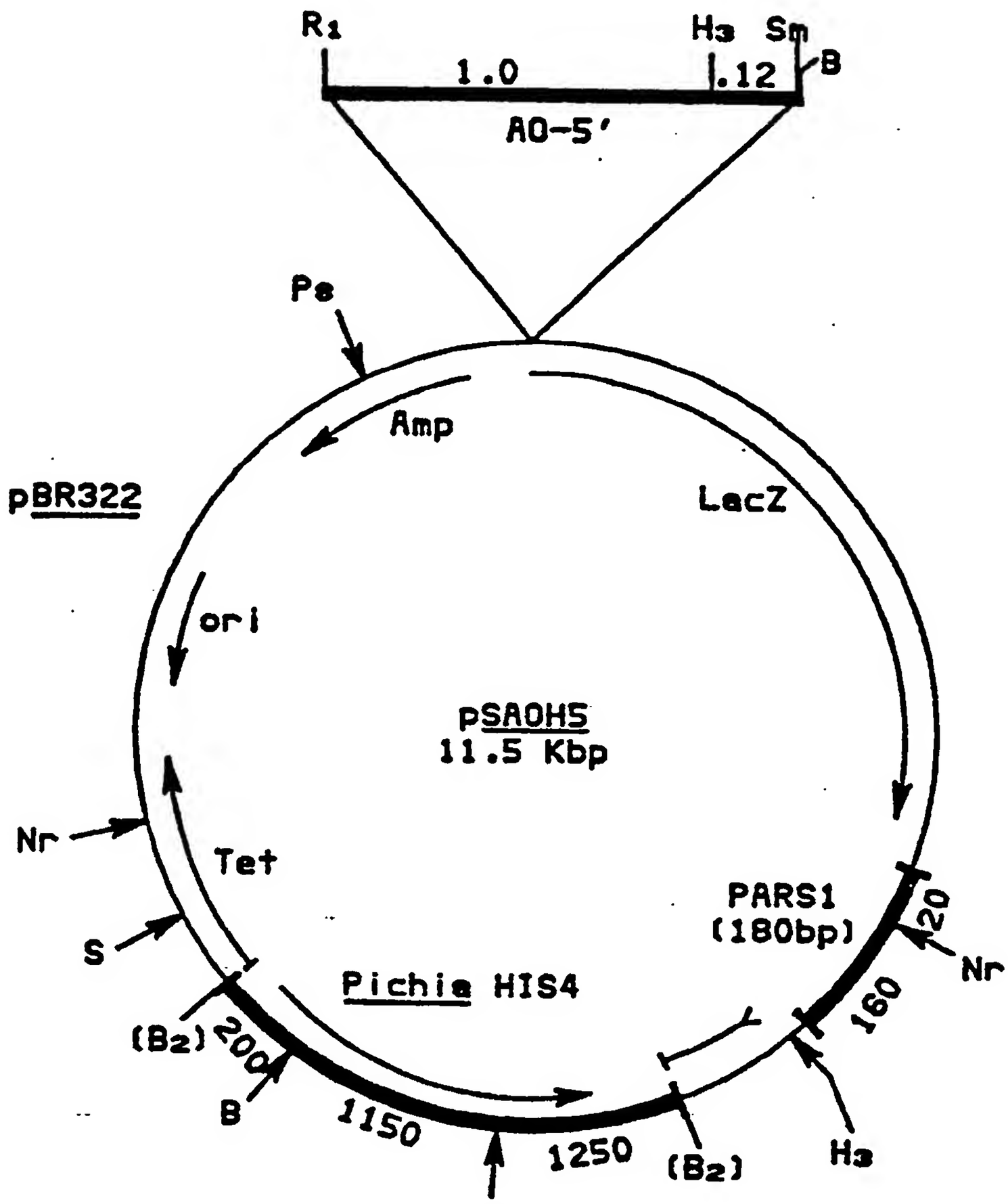
R1
H3
Sm
B
1.0
.12
AO-5'
Pe
Amp
pBR322
LacZ
ori
pSAOH5
11.5 Kbp
Nr
Tet
S
PARS1
(180bp)
20
Nr
160
Pichia HIS4
(B2)
200
B
1150
1250
(B2)
H3
S

FIG. 6